

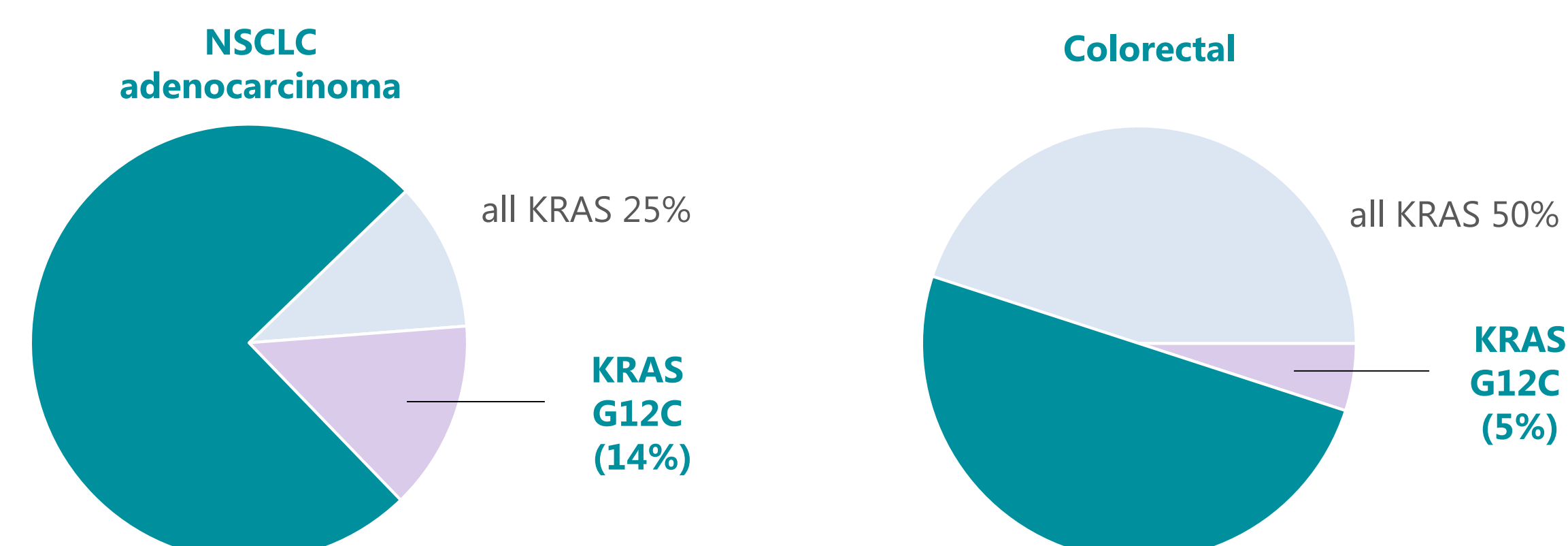
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Introduction

- KRAS is the most frequently mutated driver oncogene in human cancer¹
- The ability to target and block the function of mutated KRAS has remained elusive despite decades of research
- Findings have demonstrated that directly targeting KRAS G12C with electrophilic molecules that covalently modify the mutated codon 12 cysteine may be feasible²⁻⁶
- A novel series of potent, irreversible covalent inhibitors of KRAS G12C has been identified
- Displacement of a bound water in KRAS G12C provided a boost in activity
- Inhibitor **11** exhibited dose ascending oral bioavailability and robust in vivo target engagement
- Tumor growth inhibition in a MIA PaCa xenograft was demonstrated

KRAS G12C Mutations



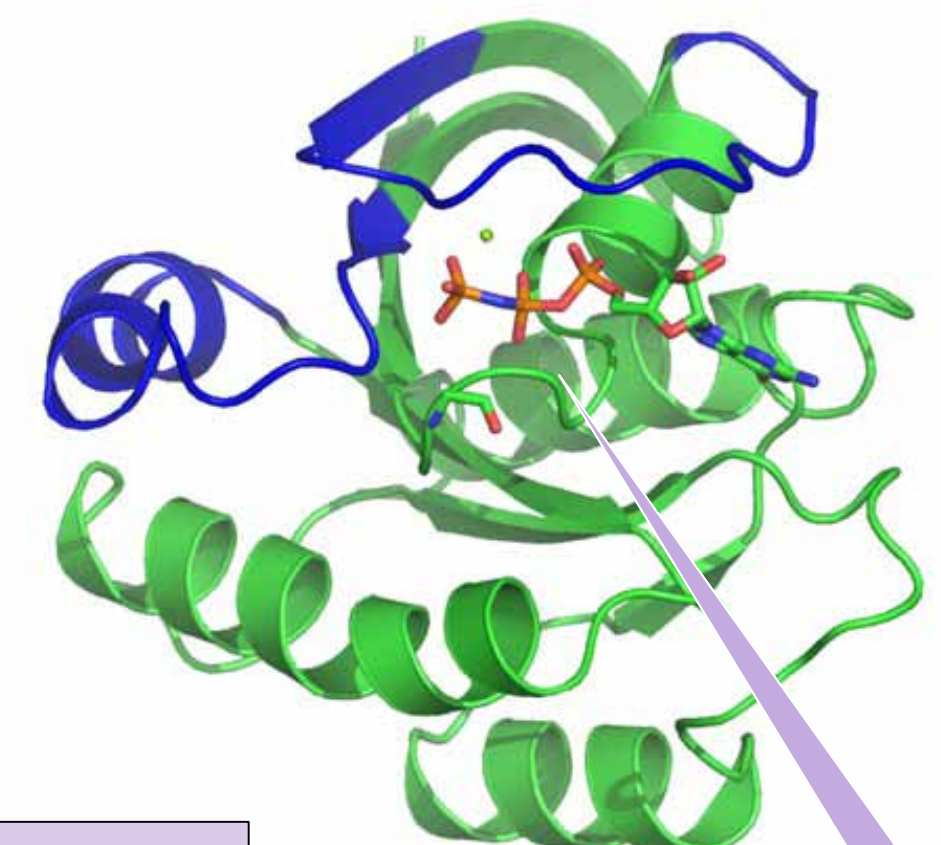
Historical Challenges in Targeting the KRAS Pathway

Upstream Inhibitors

Blocking Ras Membrane localization
• Farnesyl transferase inhibitors do not block KRAS localization

Downstream Effector Inhibitors

Raf / MEK and PI3K / AKT / mTOR
• Limited effectiveness in KRAS^{mut} tumors
• Incomplete inhibition of KRAS^{mut}
• Inhibition of KRAS^{wt} resulting in low therapeutic index



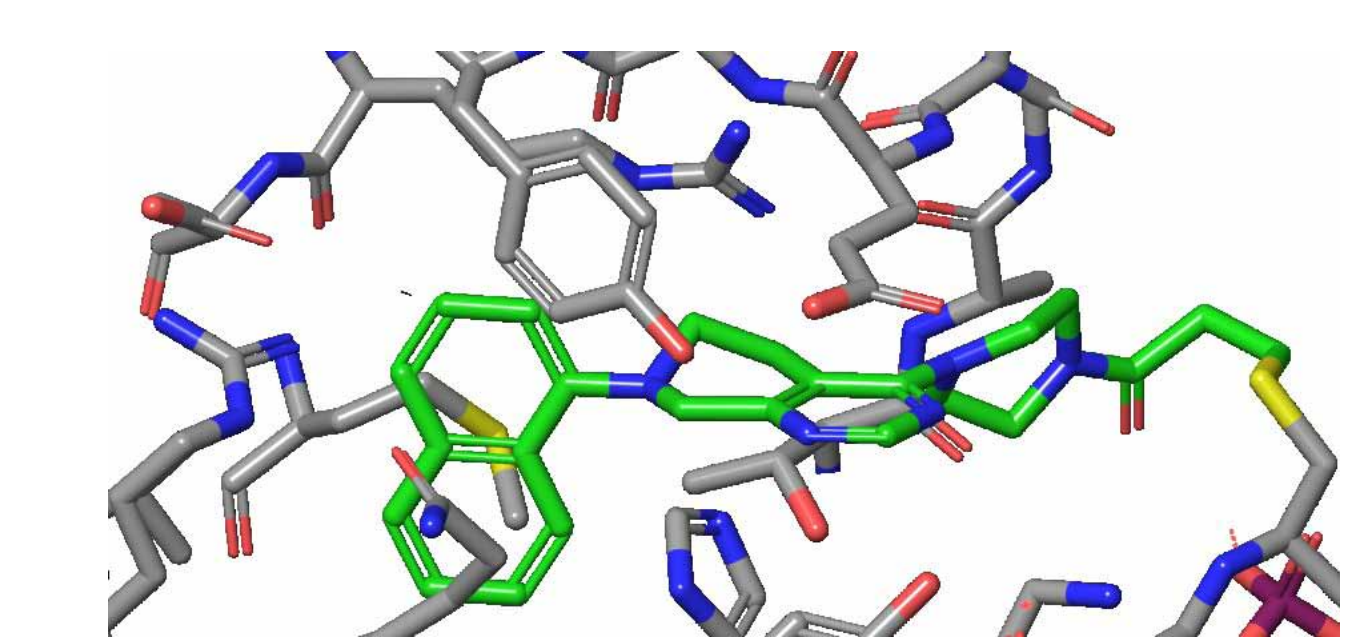
Reversible Inhibitors

Targeting KRAS^{mut} is challenging due to small, undefined catalytic site and high affinity for GTP

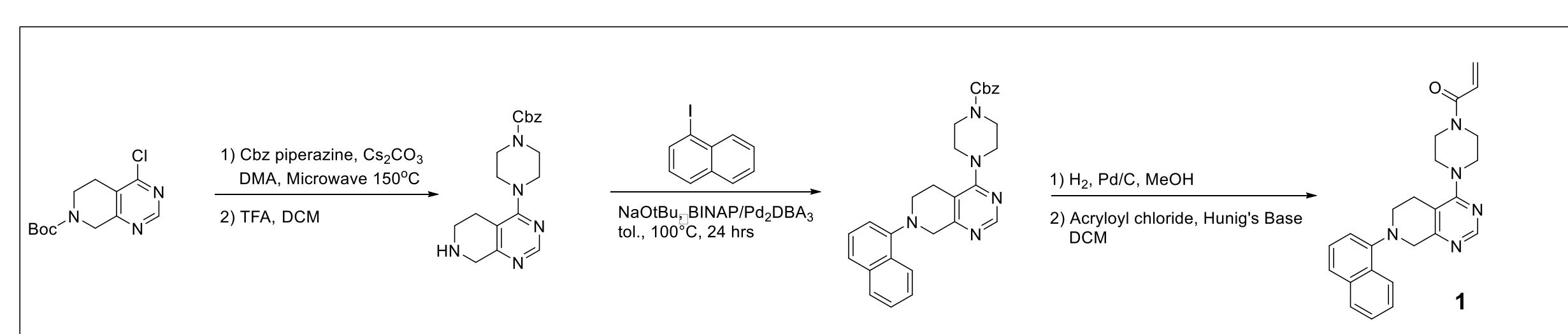
Covalent Inhibition of KRAS G12C

- Binding in the switch II pocket of GDP KRAS
- Covalent bond to cysteine 12
- Locked in the inactive conformation

Tetrahydropyridopyrimidine **1** Modifies KRAS G12C

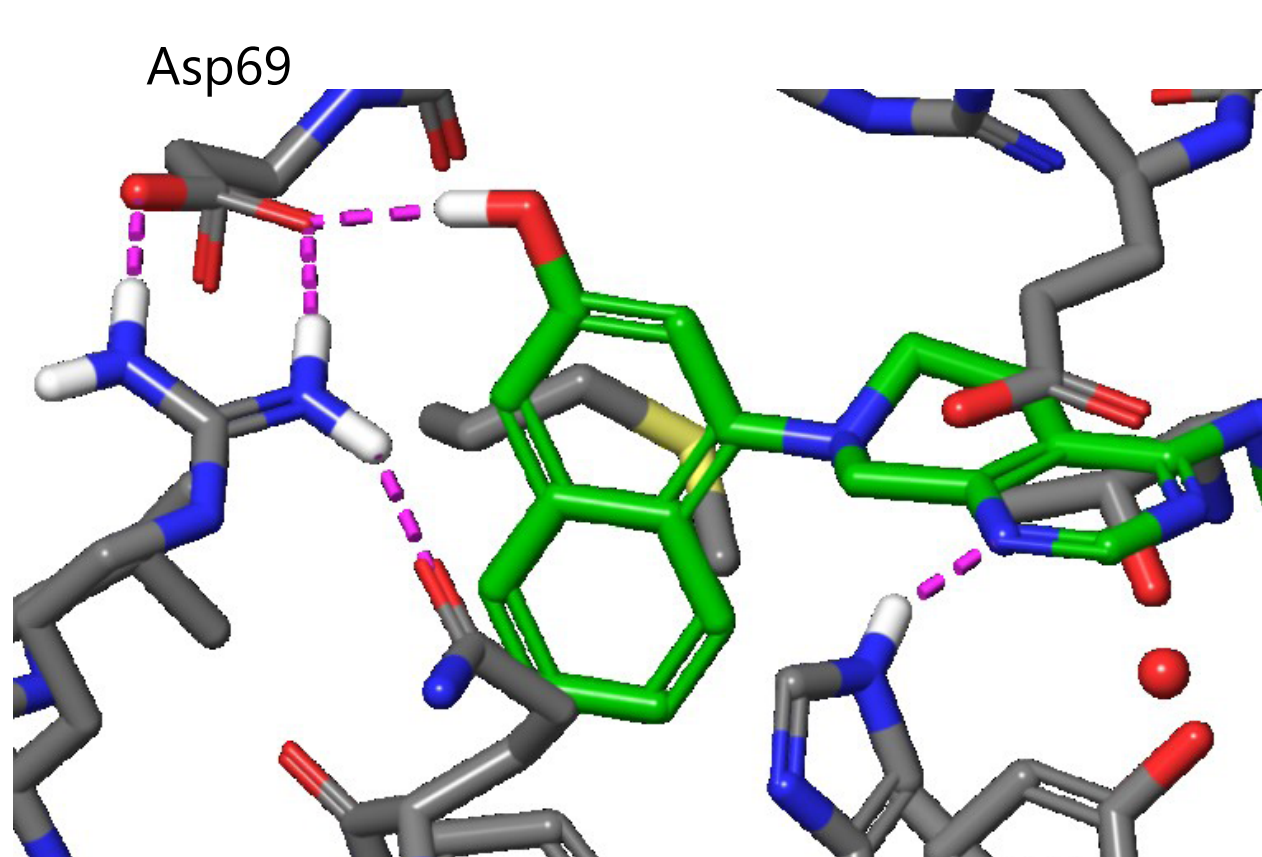


Crystal structure of **1** bound to KRAS G12C

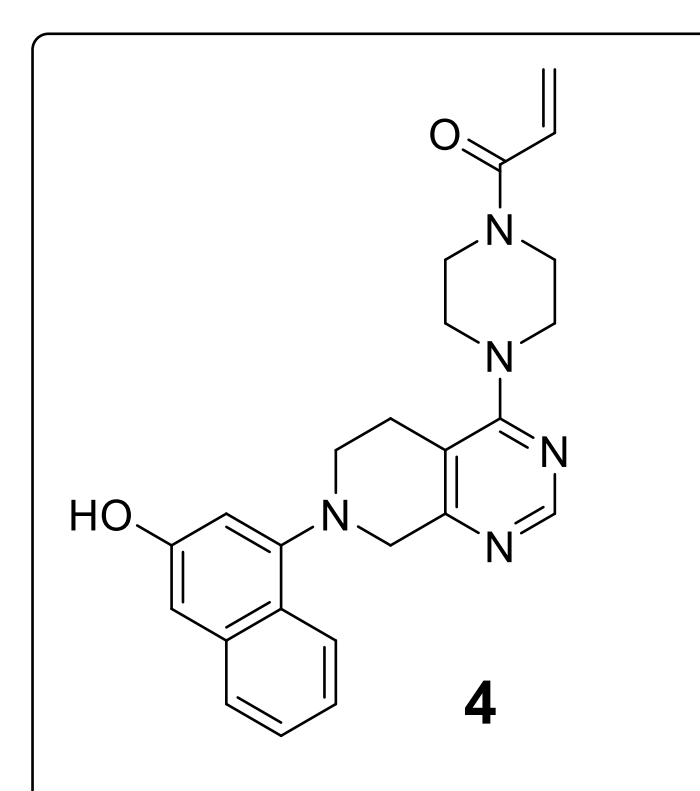


- The prototype tetrahydropyridopyrimidine **1** was synthesized in 5 steps
- **1** showed 13% modification in a 3hr/25 M protein modification assay
- This compound did not inhibit Erk phosphorylation in an H358 cell assay
- The crystal structure of **1** shows the covalent bond between the acrylamide and cysteine 12

Naphthol Makes a Hydrogen Bond to Asp69



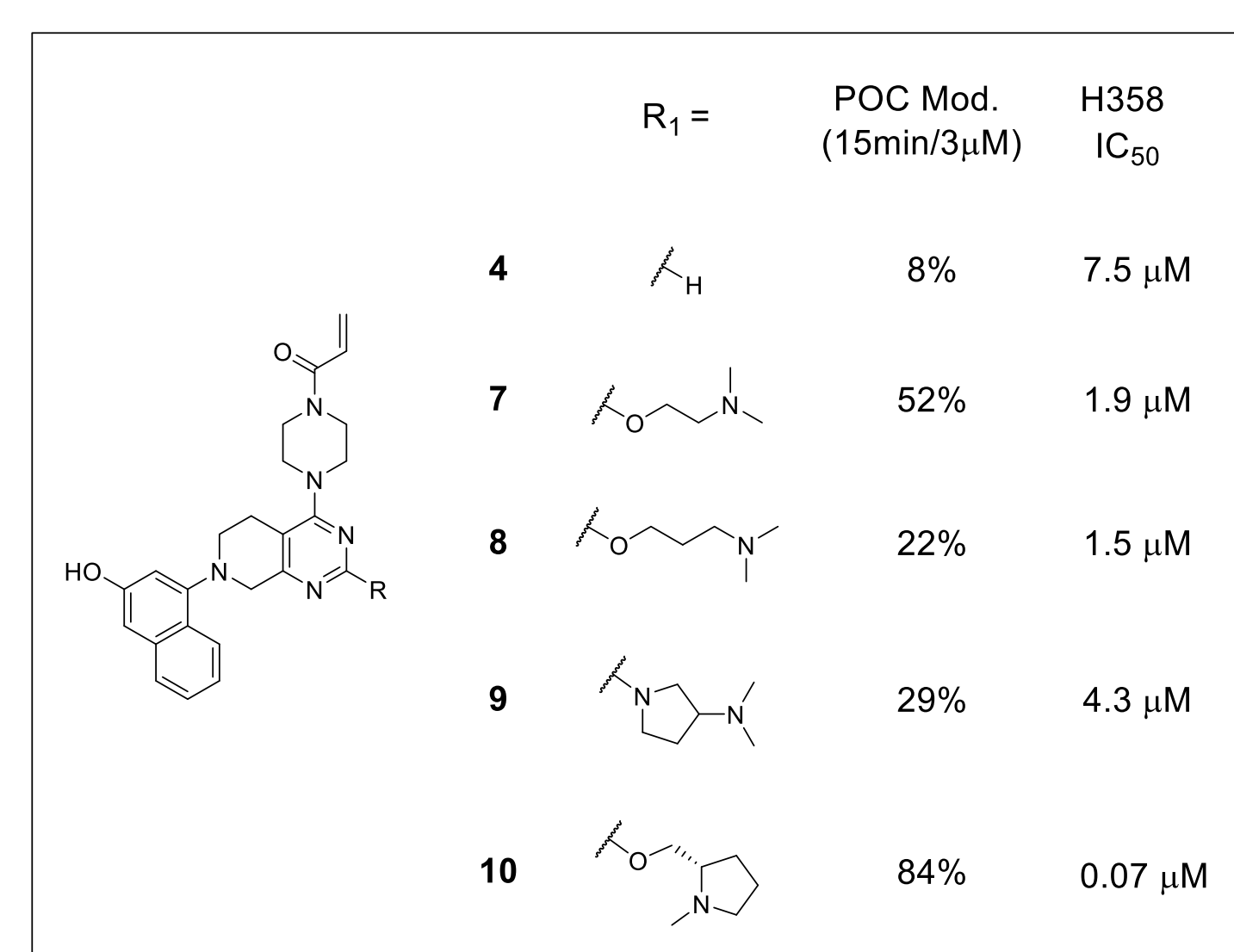
Crystal structure of **4** bound to KRAS G12C



	R ₁ =	POC Mod. (3hr/5μM)	H358 IC ₅₀ μM	R ₁ =	POC Mod. (3hr/5μM)	H358 IC ₅₀ μM
		2%	>16		99%	7.6
		22%	>16		0%	>16
		2%	>16		2%	>16

- A series of analogs were synthesized in an attempt to pick up an interaction with Asp69
- Compound **4** exhibited full modification of KRAS and inhibited Erk phosphorylation in a cell, IC₅₀ = 7.6 μM
- An X-ray crystal structure confirmed a 2.7Å H-bond between the naphthol OH and Asp69

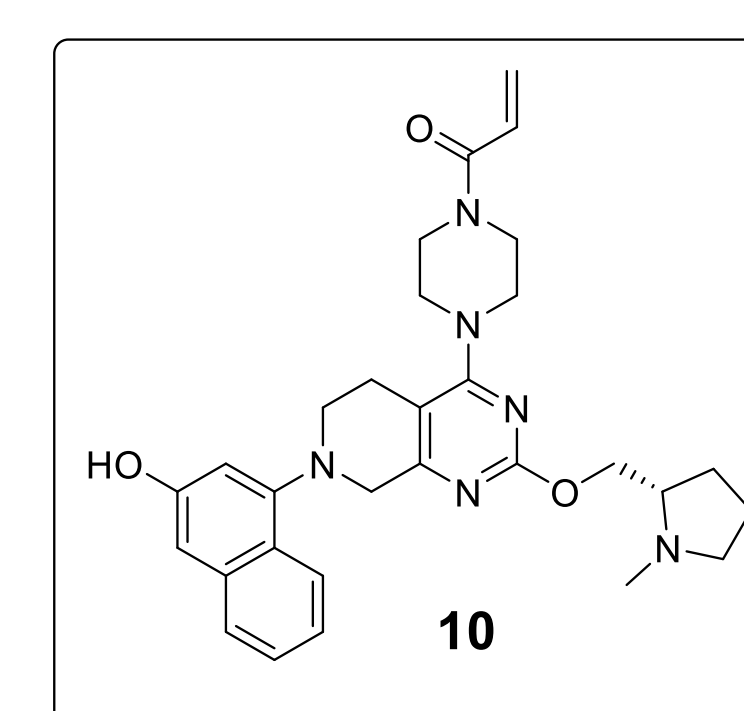
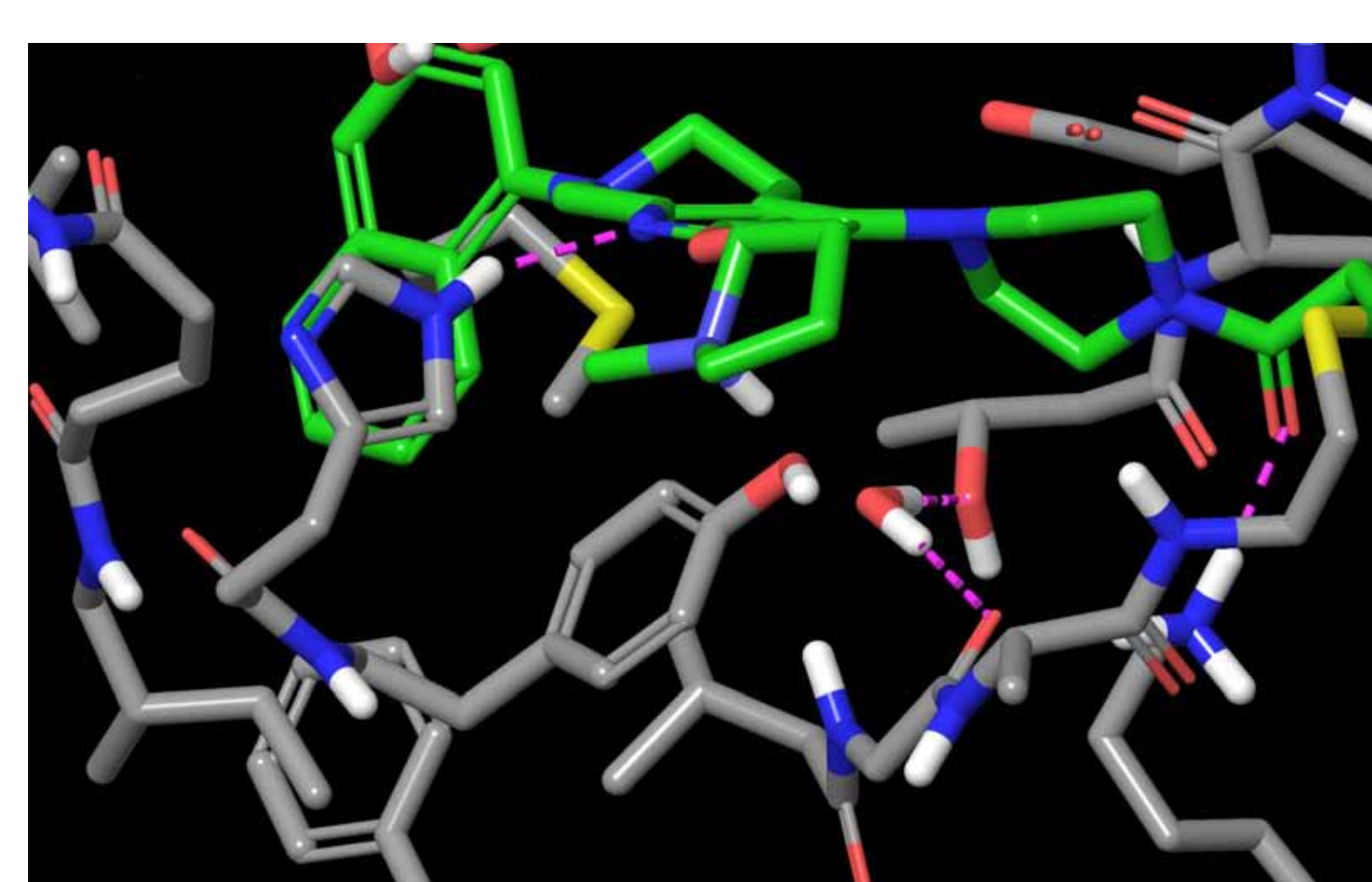
C2 Prolinol Side Chain Increases Activity



10 Mouse PK				
IV Dose	AUC _{inf} (hr ² μg/mL)	t _{1/2} (hr)	Clearance (mL/min/kg)	V _{ss} (L/kg)
3 mpk IV	1.1	0.96	46	0.53
PO Dose				
PO Dose	AUC _{inf} (hr ² μg/mL)	C _{max} (μg/mL)	T _{max} (hr)	F (%)
100 mpk	0.88	0.59	1.0	2.4

- Substitution at C2 provided compound **10** containing an N-methyl pyrrolidine side chain
- Lipophilic contacts and a hydrogen bond to Glu62 contributed to a 100x potency boost compared to **4**
- This analog inhibited KRAS dependent Erk phosphorylation with a cellular IC₅₀ = 0.07 M
- Dosed in mice, **10** showed 51% extraction ratio and oral bioavailability, F = 2.4%

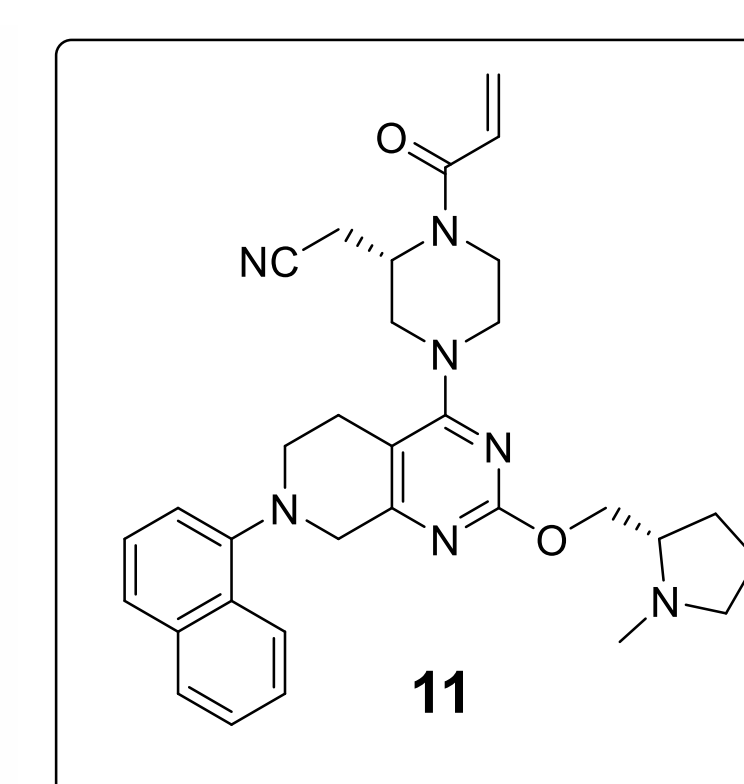
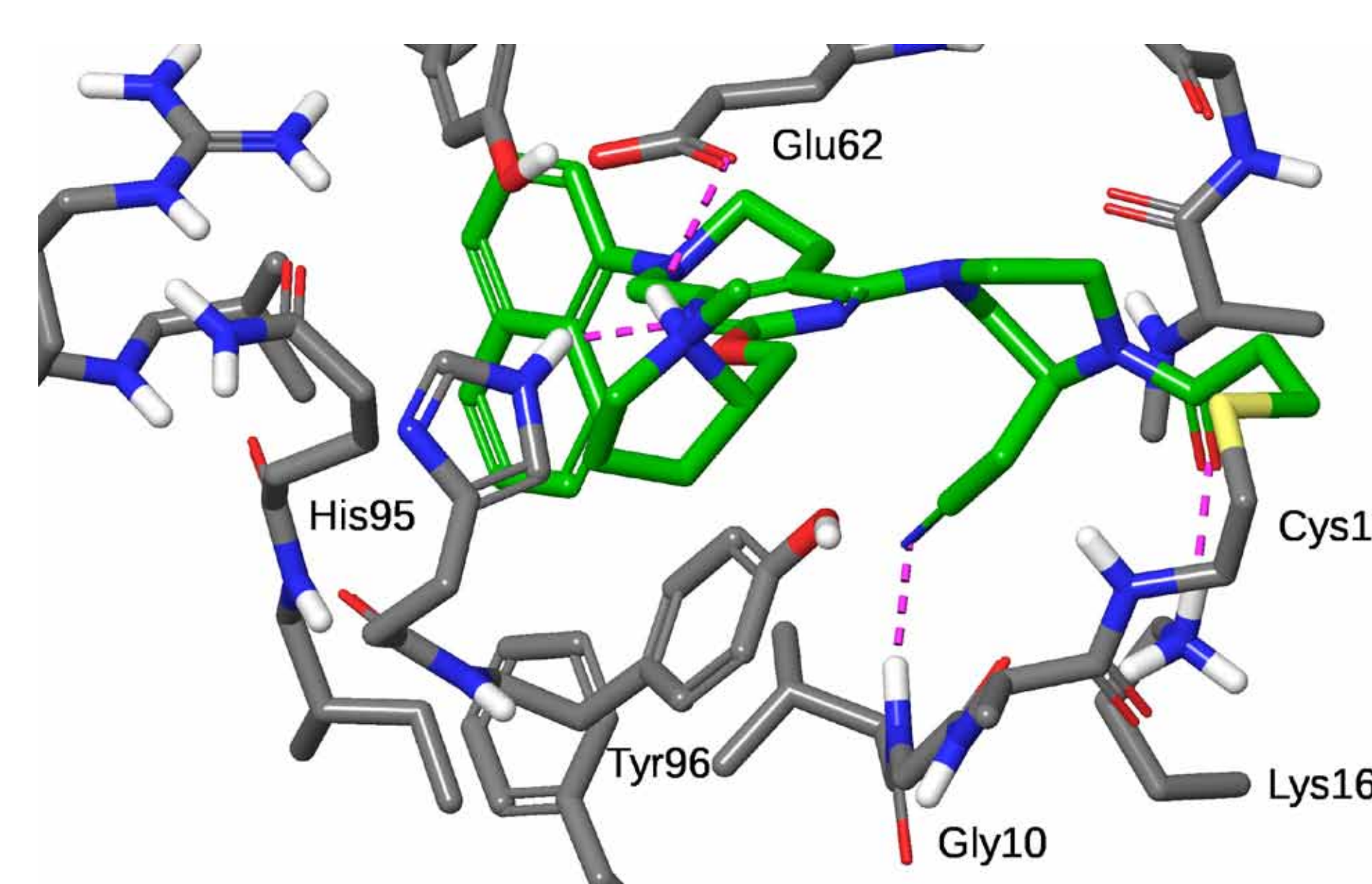
Pursue Displacement of Bound Water to Increase Activity



Crystal structure of **10** bound to KRAS G12C

- In the crystal structure of **10**, we observed a bound water complexed to Gly10 and Thr58
- It is possible that displacement of this water could lead to a potency boost⁷
- We hypothesized that removal of the hydroxy with concurrent targeting of the bound water could give a compound with desirable properties

Nitrile **11** Displaces a Bound Water in KRAS G12C

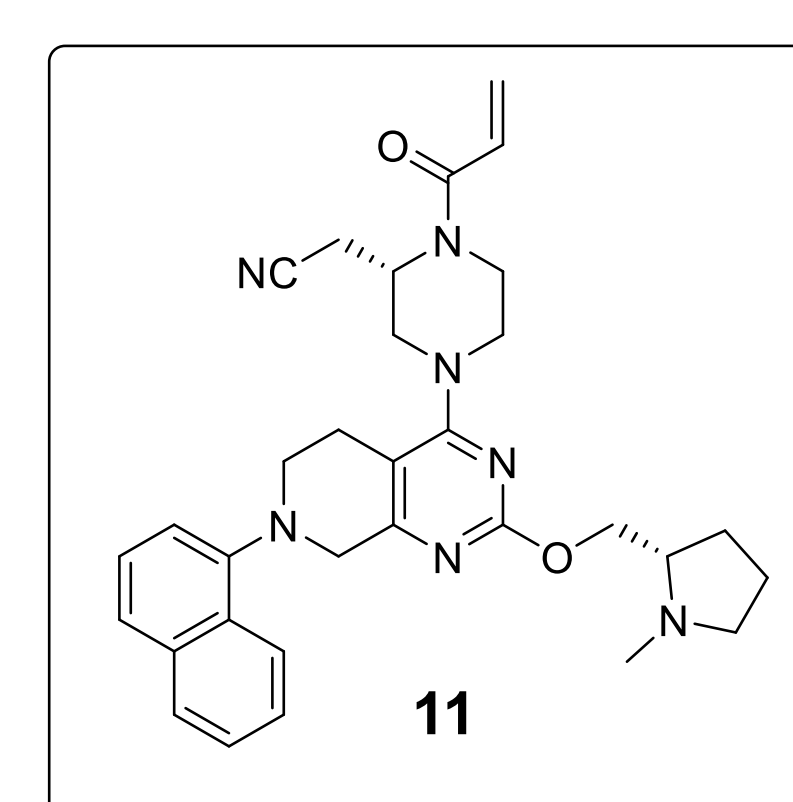


Crystal structure of **11** bound to KRAS G12C

Key Interactions

- His95 - H-bond to pyrimidine nitrogen
- Tyr96 - pi-pi interaction to the pyrimidine
- Gly10 - Nitrile H-Bond to backbone N-H
- Lys16 - Acrylamide carbonyl interaction with NH₃⁺
- Cys12 - Covalent bond formed to acrylamide olefin
- Glu62 - C2 Tail salt bridge to the acid

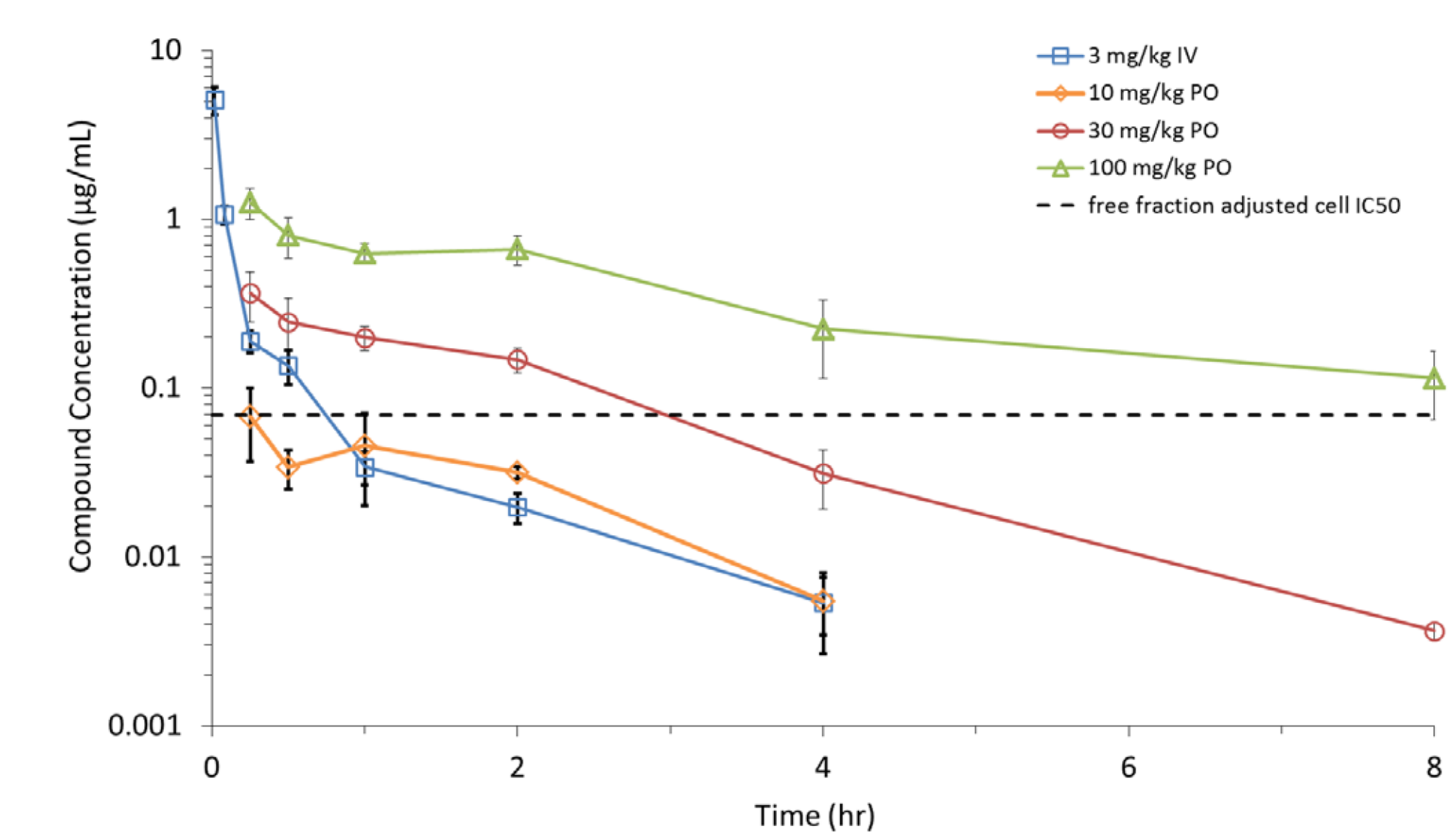
Compound **11** is an Irreversible Covalent Inhibitor of KRAS G12C



MW / ClogP / PSA	551 / 4.4 / 87
5min/3 M Protein Modification	87 %
H358 Cell IC ₅₀	5 nM
Permeability	Medium/Efflux
Mouse Hepatocyte ER	90%
Mouse Microsomal ER	49%
Mouse PPB	96%

- The inhibitor **11** showed 87% modification of KRAS G12C in the 5min/3 M modification assay
- Compound **11** inhibited KRAS dependent ERK phosphorylation in the H358 cell assay, IC₅₀ = 5 nM
- This compound has high hepatocyte ER with medium permeability and efflux
- Based on 96% protein binding the ff_{adj} cell IC₅₀ = 125 nM

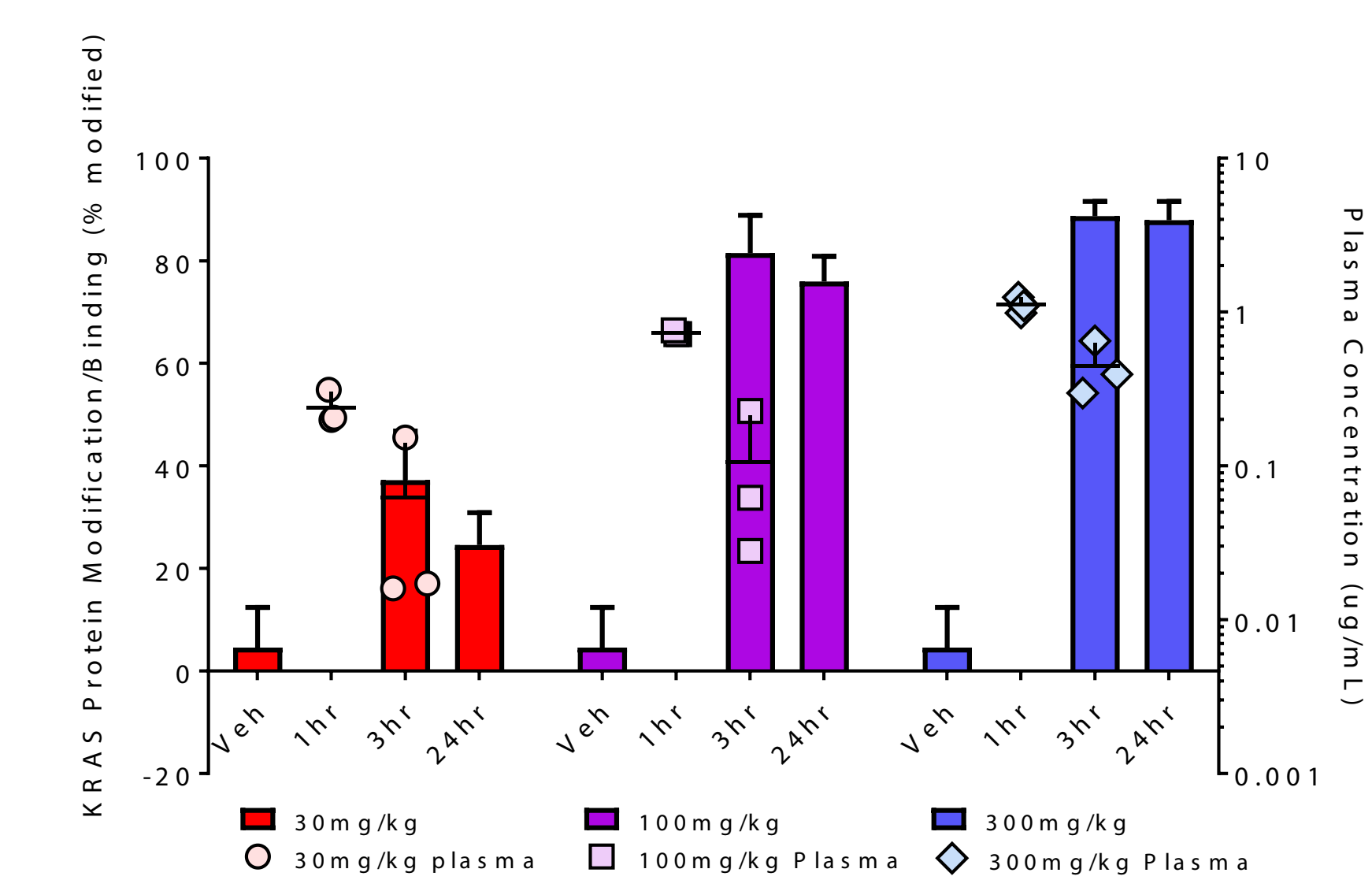
Inhibitor **11** Demonstrates Dose Ascending Oral PK in Mice



- As predicted by hepatocytes, **11** was a high clearance compound
- PO dosing resulted in super proportional increases in exposure from 10 -100 mg/kg in mice
- The 30 mg/kg dose covered the free fraction adjusted cell IC₅₀ for 3 hours

11 Mouse PK				
IV Dose	AUC _{inf} (hr ² μg/mL)	t _{1/2} (hr)	Clearance (mL/min/kg)	V _{ss} (L/kg)
3 mpk	0.50	1.1	100	2.6
PO Dose				
PO Dose	AUC _{inf} (hr ² μg/mL)	C _{max} (μg/mL)	T _{max} (hr)	F (%)
10 mpk	0.12	0.69	0.25	7.1
30 mpk	0.61	0.37	0.25	12
100 mpk	3.3	1.3	0.25	20

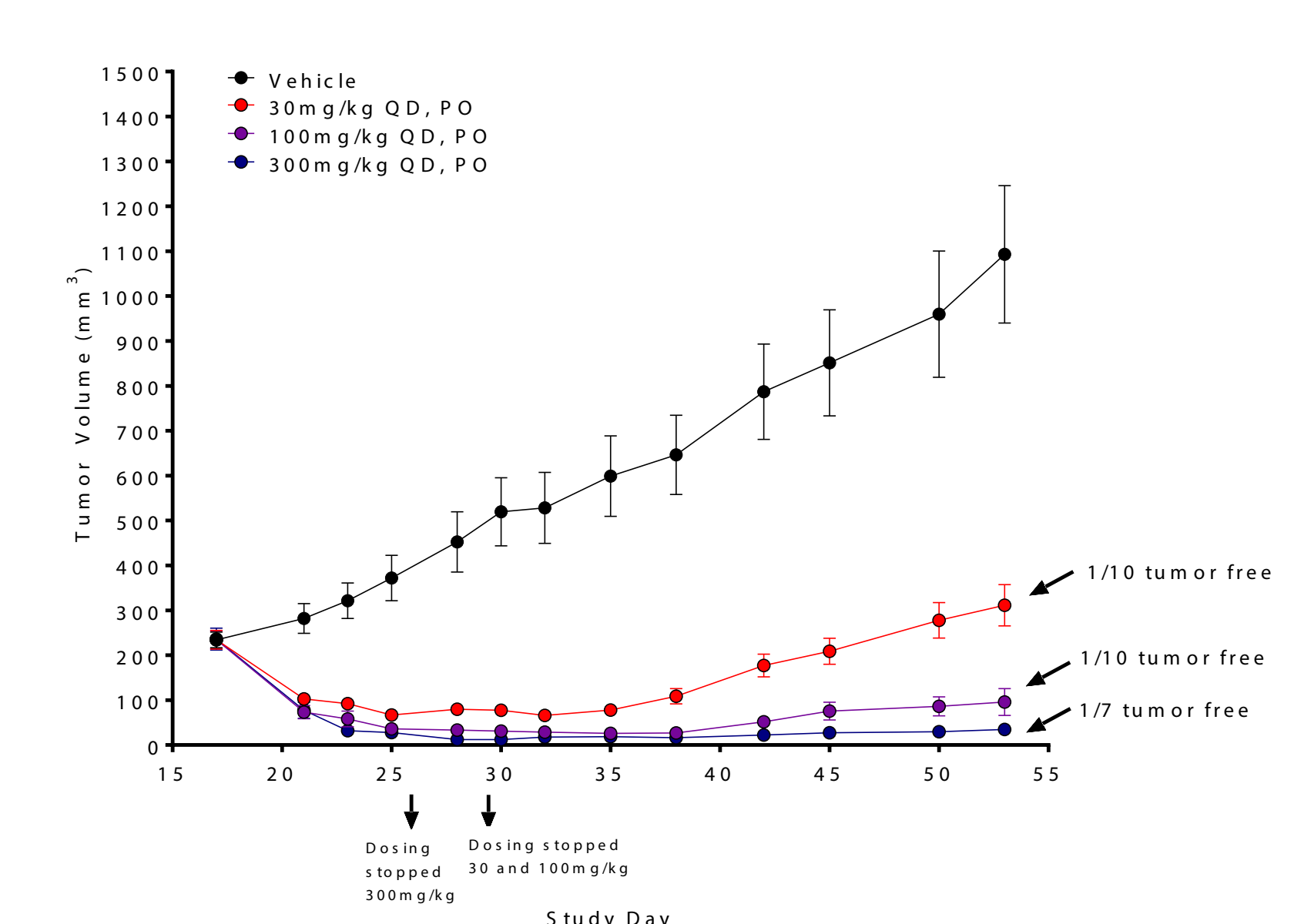
Robust Target Engagement Seen in a MIA PaCa Tumor PK/PD



- Inhibitor **11** was dosed at 30, 100, and 300 mg/kg in MIA PaCa tumor bearing mice
- Tumors were collected for processing at the 1, 3, and 24 hour time points
- Target engagement of KRAS G12C + compound was determined by mass spectrometry
- PK/PD experiments showed >80% target engagement at the 100 and 300 mg/kg dose
- This PD effect was maintained for at least 24h in this experiment

11 Exhibits Rapid and Sustained Efficacy Against MIA PaCa Xenografts

Tumor Growth Inhibition



- **11** was dosed QD PO in a MIA PaCa Antitumor Efficacy Study at 30, 100, and 300 mg/kg
- Rapid tumor growth inhibition was observed in all dose groups
- Dosing was stopped on day 26 for the 300 mg/kg group and on day 29 for the other groups
- Regrowth was observed in the 30 while the 100 and 300 mg/kg groups showed minimal regrowth
- All dose groups had animals that were tumor free at the end of the study

Summary and Conclusions

- We have identified a novel series of irreversible covalent inhibitors of KRAS G12C
- Interaction with Asp69, by hydroxy substitution of the naphthyl, led to cell activity
- 100x potency boost was gained by substitution at the pyrimidine C2 position
- Introduction of the cyanomethyl group displaced a bound water in KRAS G12C and provided an alternate path forward
- Compound **11** inhibited KRAS dependent ERK phosphorylation in the H358 cell assay with an IC₅₀ = 5 nM
- **11** demonstrated super proportional dose ascending oral exposure in mice with 3h target coverage at the 30 mg/kg dose
- In a PK/PD experiment, >80% target engagement was seen in MIA PaCa tumors with an effect lasting for at least 24 hours
- An antitumor efficacy experiment with **11**, dosed orally in mice bearing MIA PaCa tumors, showed rapid growth inhibition and durable efficacy even after dosing was stopped
- Additional compounds will be discussed in future publications

References and Acknowledgements

1. Bos, J. *Cancer Res* **49**, 4682-4689 (1989).
2. Ostrem, J. M. et al. *Nature* **503**, 548-551 (2013).
3. Lim, S. M. et al. *Angew. Chem.* **53**, 199-204 (2014).
4. Patricelli, M.P. et al. *Cancer Discovery* **6**, 317-329 (2016).
5. Ostrem, J.M., and Shokat, K.M. *Nat. Rev. Drug Discov.* **15**, 771-785 (2016).
6. Janes, M.R. et al. *Cell* **172**, 578-589 (2018).
7. Michel, J. et al. *J. Am. Chem. Soc.* **131**, 15403-15411 (2009).

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